RESEARCH

Open Access



The role of genetic testing in diagnosis and care of inherited cardiac conditions in a specialised multidisciplinary clinic

Fergus Stafford^{1,2,3†}, Neesha Krishnan^{1,2,3†}, Ebony Richardson^{1,2,3}, Alexandra Butters^{1,2,3,4}, Sophie Hespe^{2,3,4}, Charlotte Burns^{4,5}, Belinda Gray^{4,6}, Caroline Medi⁶, Natalie Nowak^{4,5}, Julia C. Isbister^{4,5,6}, Hariharan Raju^{6,7}, David Richmond^{4,6}, Mark P. Ryan⁶, Emma S. Singer^{4,5}, Raymond W. Sy^{4,5}, Laura Yeates^{1,2,3,4,5,6}, Richard D. Bagnall^{4,5}, Christopher Semsarian^{4,5,6} and Jodie Ingles^{1,2,3,5,6*}

Abstract

Background: The diagnostic yield of genetic testing for inherited cardiac diseases is up to 40% and is primarily indicated for screening of at-risk relatives. Here, we evaluate the role of genomics in diagnosis and management among consecutive individuals attending a specialised clinic and identify those with the highest likelihood of having a monogenic disease.

Methods: A retrospective audit of 1697 consecutive, unrelated probands referred to a specialised, multidisciplinary clinic between 2002 and 2020 was performed. A concordant clinical and genetic diagnosis was considered solved. Cases were classified as likely monogenic based on a score comprising a positive family history, young age at onset, and severe phenotype, whereas low-scoring cases were considered to have a likely complex aetiology. The impact of a genetic diagnosis was evaluated.

Results: A total of 888 probands fulfilled the inclusion criteria, and genetic testing identified likely pathogenic or pathogenic (LP/P) variants in 330 individuals (37%) and suspicious variants of uncertain significance (VUS) in 73 (8%). Research-focused efforts identified 46 (5%) variants, missed by conventional genetic testing. Where a variant was identified, this changed or clarified the final diagnosis in a clinically useful way for 51 (13%). The yield of suspicious VUS across ancestry groups ranged from 15 to 20%, compared to only 10% among Europeans. Even when the clinical diagnosis was uncertain, those with the most monogenic disease features had the greatest diagnostic yield from genetic testing.

Conclusions: Research-focused efforts can increase the diagnostic yield by up to 5%. Where a variant is identified, this will have clinical utility beyond family screening in 13%. We demonstrate the value of genomics in reaching an overall diagnosis and highlight inequities based on ancestry. Acknowledging our incomplete understanding of disease phenotypes, we propose a framework for prioritising likely monogenic cases to solve their underlying cause of disease.

[†]Fergus Stafford and Neesha Krishnan contributed equally to this work.

*Correspondence: jodie.ingles@populationgenomics.org.au

² Centre for Population Genomics, Garvan Institute of Medical Research, and UNSW Sydney, 384 Victoria Street, Darlinghurst, NSW 2010, Australia Full list of author information is available at the end of the article



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/ficenses/by/A/J. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Keywords: Inherited cardiac conditions, Sudden cardiac death, Genetic testing

Background

Inherited cardiac conditions are associated with significant morbidity and mortality and can result in sudden cardiac death (SCD) at a young age. Clinical genetic testing is a key aspect of clinical management [1], currently used primarily for cascade testing of asymptomatic atrisk relatives. For some, a genetic result can also change diagnosis, clarify prognosis, and guide clinical management. However, the diagnostic yield of cardiac genetic testing remains low, with only ~ 40% having a likely pathogenic or pathogenic variant (LP/P) identified [2, 3]. Critical gaps remain in our understanding of the phenotypic spectrum and genetic architecture of these diseases, including growing acknowledgement of a role for complex polygenic contributions to disease [4, 5].

Efforts to improve the diagnostic yield of genetic testing thus far have largely focused on patients who fulfil the clinical diagnostic criteria for an inherited cardiac condition. It is unclear what proportion have undifferentiated cardiac pathology that does not meet widely accepted specific clinical diagnostic criteria, i.e. undiagnosed cardiac disease. The role genetic testing can play in this setting is far less clear but presents an exciting opportunity to improve the diagnostic yield through gene discovery and mechanistic insights, particularly among those with features suggestive of a monogenic cardiac disease.

Individuals with monogenic diseases are more likely to have a younger age at onset and a positive family history, compared to their gene-negative counterparts [6-8]. Recent studies have shown HCM patients with causative variants in sarcomere genes have more severe disease and worse outcomes compared to sarcomerenegative patients [6, 9], and more generally, individuals with phenotypic extremes are enriched for rare, large effect size variants [10, 11]. Determining the likelihood of monogenic disease could provide a powerful tool in discriminating between monogenic and complex disease aetiology and allow prioritisation of cases where additional gene discovery efforts will be of greatest utility.

Here, we describe our experience of evaluation and diagnosis of individuals seen in a specialised multidisciplinary cardiac genetic clinic over an 18-year period. These clinics have largely included clinical and genetic reviews of families with inherited cardiomyopathies, arrhythmia syndromes, and evaluation after a young SCD. Those with a concordant clinical and genetic diagnosis were considered solved, and we report the proportion in which the final diagnosis was clarified or changed due to genetic testing. We describe the role of genomics in elucidating an overall diagnosis and propose a simple scoring system for prioritising difficult unsolved cases most likely to have a monogenic disease basis. Clarifying the underlying diagnosis in these unsolved cases may provide important insights about disease mechanisms and lead to an increased yield of cardiac genetic testing with the potential for important clinical and prognostic implications worldwide.

Methods

Study population

Consecutive, unrelated probands referred to a specialised, multidisciplinary clinic at Royal Prince Alfred Hospital (Sydney, Australia) between 2002 and 2020 were evaluated. These clinics were initially established for the management of hypertrophic cardiomyopathy (HCM); however, since 2003, they have expanded to include inherited cardiomyopathies, arrhythmia syndromes, and sudden cardiac death evaluation [12]. Patients were included if they were the proband, i.e. the first affected person in the family to attend the clinic. Probands were included if they were seen in the clinic, had definite or probable cardiac disease, and underwent comprehensive clinical investigation and genetic testing. Families where the only affected family member was deceased and not evaluated during life were excluded. Ethics approval was granted by the Sydney Local Health District Human Research Ethics Committee (X20-0522 and 2020/ETH03142) including a waiver of consent to collect a minimum de-identified dataset.

Clinical investigations and baseline diagnosis *Cardiac evaluation*

Individuals were evaluated per clinical recommendations at the time of consultation. Most patients underwent transthoracic echocardiography and 12-lead electrocardiography (ECG) in clinics as a minimum. Where this was not possible, the results of previous investigations were requested from the referring clinician or hospital. Further investigations were conducted at the discretion of the treating cardiologist and included the following: modified 'high' right precordial ECG (V1 and V2 cranially displaced into 2nd and 3rd intercostal spaces), 24-h ambulatory Holter monitoring (including 12-lead and/ or modified high right precordial lead positions), exercise stress testing, signal-averaged ECG, and drug provocation tests (including flecainide or ajmaline challenge depending on the year performed). Cardiac magnetic resonance (CMR) imaging was routinely performed to

identify the structural changes including myocardial non-compaction where echocardiography was negative or inconclusive and when assessing for arrhythmogenic cardiomyopathy (ACM). Cardiac investigations were reported by cardiologists as per clinical practice and according to the accepted guidelines [13–15].

Family history

Family history was evaluated by experienced cardiac genetic counsellors from 2003 and included a 3-generation pedigree identifying affected relatives and any significant cardiac events such as SCD and cardiac transplant. Details were confirmed with medical records and postmortem reports where available.

Baseline phenotype and clinical diagnosis

The clinical diagnosis after baseline evaluation was classified as definite or uncertain. A definite clinical diagnosis included any individual meeting diagnostic criteria, with or without atypical features. An uncertain clinical diagnosis included those with an abnormal but uncertain phenotype, such as possible (borderline) diagnoses and undiagnosed (unclassified) cardiac disease. Cases were adjudicated to exclude those with likely physiological changes and discussed in regular multidisciplinary meetings with other clinical team members as needed.

Time to diagnosis

For those solved using additional genetic testing research efforts, time to diagnosis in years from the first presentation in the clinic to a genetic diagnosis was ascertained. Time to diagnosis was calculated within two decades: 2002–2010 and 2011–2020.

Genetic testing and variant interpretation

Cardiac genetic testing including targeted gene panels, whole-exome sequencing and whole-genome sequencing was performed clinically (i.e. in a clinically accredited laboratory) or on a research basis and as per best practice at the time of sequencing. Approaches to sequencing evolved significantly over time. Variants were assessed and classified using MYH7-modified American College of Medical Genetics and Genomics and Association of Molecular Pathologists (ACMG/AMP) standards for sequence variant interpretation [16, 17]. Likely pathogenic and pathogenic (LP/P) variants were considered causative/solved. Where no variants were identified, or were classified as benign, likely benign, or non-suspicious VUS, genetic testing was considered to be negative, i.e. unsolved. Patients with variants of uncertain significance (VUS) deemed suspicious but lacking critical evidence were not considered unsolved and are shown separately. Suspicious VUS were absent or very rare, i.e. < 0.004% for hypertrophic cardiomyopathy (HCM) and more stringent for other less prevalent diseases [18], in population databases (Genome Aggregation Database; gnomAD), [19] occurred in a gene with definitive gene-disease association relevant to the baseline phenotype, and predicted to alter the protein sequence. In addition, suspicious VUS had some evidence for causation but not enough to reach the LP/P classification. Genes with a robust gene-disease association at the time of interpretation were considered tier 1 genes, while tier 2 genes were those with less wellcharacterised clinical validity (Additional file 1: Table S1) [20–23]. Gene panels were mostly analysed, and in some, expanded genetic testing was performed which involved the analysis of ~ 500 cardiac genes as part of research studies (tier 2 analysis). Mitochondrial DNA variants were not routinely evaluated, except for those who underwent whole-genome sequencing as part of gene discovery studies.

Likelihood of monogenic disease

Based on the available literature and our experiences in the clinical setting, we considered a severe phenotype, young age of disease onset, and positive family history as characteristics more likely to be associated with monogenic disease, as opposed to a complex disease arising from environmental factors and/or the accumulation of common genetic variants with small effect sizes. The scoring system and disease-specific high-risk/severe phenotype definitions are shown in Fig. 1A, incorporating the characteristics from the family unit. Familial disease included > 1 relative with a phenotype consistent with an inherited cardiac disease. Severe phenotype included ventricular arrhythmia (SCD, resuscitated cardiac arrest, sustained VT, appropriate shock for VT/VF by implantable cardioverter defibrillator); heart failure (cardiac transplant, left ventricular assist device, ejection fraction < 30%); and phenotype extremes (e.g. left ventricular hypertrophy [LVH] \geq 30 mm, QTc \geq 500 ms; Additional file 1: Table S4). Young age included those < 30 years (1) point) or < 10 years (2 points). A higher score indicated a greater likelihood of monogenic disease.

Final diagnosis and clinical impact

Solved cases were those where the clinical diagnosis and genetic findings were concordant and adequately explained disease. When additional research was required to make a genetic diagnosis, these were considered 'solved on review', e.g. beyond standard clinical practice at the time including analysis of new genes and functional studies. Cases were considered 'unsolved' if no genetic cause for their disease was identified. If no genetic variants were identified with one or more features indicative of monogenic disease present, those



individuals were considered unsolved and more likely to have a monogenic basis ('likely monogenic'). Conversely, individuals with no causative genetic variant and without these features were considered unsolved and more likely to have a complex aetiology ('likely complex').

Among solved cases, we evaluated whether the genetic result clarified inheritance risk, family screening recommendations, frequency and type of cardiac investigations, additional specialist consultations (e.g. neurological review), changes to medication, and lifestyle advice arising from their genetic diagnosis.

Ancestry

Self-reported ancestry was recorded for most probands and classified according to the Australian Standard Classification of Cultural and Ethnic Groups [26]. Ancestry groups included European, East Asian, South and Central Asian, Middle-Eastern and North African, or other ancestries (including less frequent ancestries such as Sub-Saharan African, Oceanian, and People of the Americas). We compared the proportion of patients with LP/P variants, suspicious VUS, and unsolved between all groups to assess the diagnostic yield. Representation of ancestry groups was compared to the Greater Sydney population using the Australian Bureau of Statistics 2016 Census data. Census data include a proportion who identify as 'Australian', who have typically included a high number of Anglo-Celtic individuals, and who were considered to have European ancestry.

Statistical analysis

Data were analysed using R (version 4.1.0) and Graph-Pad Prism (version 9.2.0). Descriptive statistics were performed, including the chi-squared tests for categorical variables and Student's *t*-test for continuous variables. A *p*-value of < 0.05 was considered statistically significant. R packages *ggplot2* and *ggalluvial* were used for figures.

Results

Study population and baseline clinical diagnoses

There were 1697 probands who attended the specialised clinic between 2002 and 2020, and 888 met the inclusion criteria, while 809 were excluded due to no genetic testing performed (348, 43%), no living affected family member seen in the clinic (318, 39%), cardiac findings determined to be physiological (e.g. hypertensive heart disease, athletes' heart; 136, 17%), or limited information or clinical evaluation not performed (7, 1%; Fig. 2). Genetic testing was not performed for several reasons, including inability to provide funded clinical testing at times, not meeting the criteria for research studies including genetic testing, or patient declined. Of the 888,



Table 1 Clinical characteristics of included probands

	Definite diagnosis	Uncertain clinical diagnosis	Total cohort
Age, years	56.0 ± 16.5	46.1 ± 15.9	54.8 ± 16.7
Age at diagnosis, years	42.3 ± 16.6	36.5 ± 16.1	41.6 ± 16.6
Male sex	487 (64)	71 (56)	558 (63)
Severe phenotype	209 (27)	65 (52)	274 (31)
Sudden cardiac events	84 (11)	51 (40)	135 (15)
Heart failure	7 (< 1)	0 (0)	7 (< 1)
Transplantation	9 (1)	3 (< 1)	12 (14)
Phenotype extremes	74 (10)	8 (6)	82 (9)
Ventricular arrhythmia	30 (4)	3 (2)	33 (4)
Positive family history	247 (32)	36 (29)	283 (32)
Age at diagnosis ≤ 30 years	192 (25)	52 (41)	244 (27)
Age at diagnosis \leq 10 years	7 (< 1)	0 (0)	7 (< 1)
Inherited cardiomyopathy	662 (87)	62 (49)	724 (82)
Inherited arrhythmia syndrome	100 (13)	24 (19)	124 (14)
Undiagnosed	0 (0)	40 (32)	40 (5)
Total	762 (86)	126 (14)	888 (100)

Data are shown as mean \pm standard deviation or *n* (%)

a genetic cause of disease was identified in 403 (45%), while 485 (55%) remained unsolved.

The demographic and clinical characteristics are summarised in Table 1. Following baseline evaluation, a definite clinical diagnosis was made in 762 (86%) of the total cohort. The remaining 126 (14%) had uncertain clinical diagnoses, including 21 (17%; 2% of the total audit cohort) with undiagnosed primary cardiac phenotypes, 36 (28%) with unexplained resuscitated sudden cardiac arrest (SCA), and 69 (55%) with possible disease. The majority of those with a definite clinical diagnosis at baseline were diagnosed with HCM (589/762, 77%), Brugada syndrome (n = 50; 7%), or long QT syndrome (n =44; 6%). Less common diagnoses, accounting for 1-5%of those with a definite diagnosis, included dilated cardiomyopathy (DCM), left ventricular non-compaction (LVNC), arrhythmogenic cardiomyopathy (ACM), and catecholaminergic polymorphic ventricular tachycardia (CPVT). Noonan syndrome (with LVH), restrictive cardiomyopathy, and sick sinus syndrome each accounted for < 1%.

Genetic findings

Genetic testing identified LP/P variants in 330 (37%) of the total cohort and suspicious VUS in 73 (8%), and the remainder had no variants of interest (n = 485; 55%; Fig. 3A). There were 209 distinct variants reported and classified as LP/P (Additional file 1: Table S2) and 73 as suspicious VUS (Additional file 1: Table S3). The yield of

LP/P variants and suspicious VUS was similar for those with inherited cardiomyopathies and inherited arrhythmia syndromes (Fig. 3B).

Variants classified as suspicious VUS (n = 73) were most commonly identified in MYBPC3 (17/73, 23%), followed by *MYH7* (n = 15, 21%), *TNNI3* (n = 6, 8%), TNNT2 (n = 5, 7%), KCNH2 (n = 4, 5%), FLNC (n = 4, 5%)4, 5%), and *PRKAG2* (n = 3, 4%). While these variants lacked critical evidence supporting pathogenicity, existing evidence included rarity in the general population (PM2; n = 73, 100%), in silico tools supporting a deleterious effect (PP3; n = 57, 78%), enrichment in cases compared to controls as a supporting (PS4 supporting; n =17, 23%) or moderate (PS4 moderate; n = 5, 7%) level of evidence, whether the variant was located in a hotspot region (PM1; *n* = 5; 7%; and PM1_supporting *n* = 2, 3%), and segregation with disease in a family (PP1, co-segregation with greater than three meioses; n = 4; 6%). Other criteria applied included PM6 (assumed de novo without confirmation of paternity and maternity), PVS1_moderate (null variant in a gene where loss of function is a known disease mechanism), and PS3_supporting (wellestablished functional studies), applied once each.

Achieving a final genetic diagnosis

There were 357 (40%) of the total cohort who were considered solved using standard clinical-genetic approaches (solved at baseline) and an additional 46 (5%) solved on review following additional effort. Evaluation of tier 2



gene lists (via whole exome and genome sequencing) in individuals not solved at baseline identified LP/P variants in a further 17/370 (5%) and suspicious VUS in 7 (2%).

A genetic diagnosis clarified the clinical diagnosis for 51 (13%) of the 403 patients with an LP/P variant (Fig. 4). This included six individuals with an undiagnosed primary cardiac disease in whom genetic testing provided a diagnosis, including a *TTN* truncating variant in a patient with conscious VT, mitral valve prolapse, and mild LV dysfunction; an *NDUFB11* variant in two individuals with atypical LVH, clarifying the phenotype as histiocytoid cardiomyopathy; a missense variant in *CACNA1C* in one individual with an atypical LVH and arrhythmia phenotype; a missense *DES* variant in one individual with complete infranodal heart block and subsequent VF arrest; and an *SCN5A* truncating variant in one individual with atrial flutter and complete heart block.

Among the 46 (5%) solved on review following additional effort, a genetic diagnosis was most commonly achieved using research-focused genetic testing (Additional file 1: Fig. S1). Often multiple factors were required to achieve the final diagnosis. Time-to-diagnosis in this group was prolonged for those clinically diagnosed in the years 2002–2010 (n = 16, mean 11.9 years SD 5.7) compared with 2011–2020 (n = 30, mean 2.7 years SD 5.4; p < 0.01).

Among the cases solved on review, 20/46 (43%) had an uncertain baseline clinical diagnosis. The most common phenotype at baseline evaluation was HCM (20; 43%) or possible HCM (borderline LVH; 6, 13%). All had undergone prior genetic testing that failed to identify a cause, with reasons including genes not previously tested, variants located in deep intronic regions, or lack of available evidence for pathogenicity at the time of testing (e.g. reported proband counts, segregations). Subsequent sequencing identified six with MYBPC3 variants (4 deep intronic splice site altering, 1 synonymous affecting splicing, and 1 missense variant) [24]; four with missense variants in *FLNC*; three with truncating or canonical splice site variants in FHOD3 [25]; two with missense variants in ACTN2 [26]; two with missense variants in MYH7, one each with a missense variant in TNNI3, PRKAG2, and MT-TI; a de novo variant in DES; one individual with asymmetric LVH and high burden of ventricular ectopy with the pathogenic p.S358L variant in *TMEM43*; and one with SCD < 30 years and postmortem findings suggesting possible HCM but a family history of Brugada syndrome and sinus node disease with a balanced translocation impacting *SCN5A* [27]. Two individuals presented with LVNC, and genetic testing identified putative loss of function variants in *TBX5* with subsequent clinical review confirming a diagnosis of Holt-Oram syndrome [28].

Ancestry

Self-reported ancestry was available for 541/608 probands (89%). The majority were of European ancestry (n = 403, 75%), 40 (8%) East-Asian, 30 (6%) South Asian, 43 (8%) Middle-Eastern and North African, and 26 (5%) categorised as other (comprised Oceanian, Sub-Saharan African, and People of the Americas' ancestries).

The diagnostic yield differed based on ancestry group (Fig. 5). Middle-Eastern and North African patients had the lowest yield of LP/P variants (12/43, 28%), a high rate of suspicious VUS (n = 8, 19%), with more than half of patients having no clear genetic cause for their disease (n = 23, 54%). Conversely, the diagnostic yield was the highest among probands with South Asian (LP/P, 67%), European (LP/P, 57%), and other (LP/P, 62%) ancestry groups. When compared to Europeans (41/403, 10%), the yield of suspicious VUS was higher in other ancestry groups (15–20%; chi-squared test, p < 0.05).

Comparison with the prevalence of ancestry groups in the clinic cohort versus Greater Sydney was as follows: East Asian (7.4% versus 14.8%), South Asian (5.5% versus 6.4%), Middle Eastern and North African (7.9% versus 6.0%), and European (75.5% versus 67.9%) patients.

Impact of a genetic diagnosis

Among 26 of 46 (57%) who were solved on review, the genetic diagnosis resulted in a change in management and risk stratification for the proband and/or their family. There was a change in the inheritance risk in 11 (22%), including eight where X-linked inheritance was confirmed, two with de novo variants and one with a mitochondrial variant that confirmed maternal inheritance. Clinical screening recommendations for family members were altered in 14 (30%), such as the need for CMR imaging, drug provocation challenge, and/or referral to other (non-cardiac) specialised clinics. Additional specialist review was indicated for 16 (35%) probands, including neurologist for three (7%) with Emery-Dreifuss muscular dystrophy due to FHL1 variants previously diagnosed as HCM, three (7%) with myofibrillar myopathy due to DES variants, and two (4%) undiagnosed individuals with histiocytoid cardiomyopathy due to variants in *NDUFB11*;

(See figure on next page.)

Fig. 4 Impact of genomics in clarifying or changing the overall diagnosis. Baseline phenotype of ARVC included those who met the 2010 Task Force Criteria. *Abbreviations*: ARVC, arrhythmogenic right ventricular cardiomyopathy; DCM, dilated cardiomyopathy; HCM, hypertrophic cardiomyopathy; LQTS, long QT syndrome; LVNC, left-ventricular non-compaction; CPVT, catecholaminergic polymorphic ventricular tachycardia; ACM, arrhythmogenic cardiomyopathy

ARVC		НСМ
		LQTS
DCM		ACM
		ACTIVE cardiorityopatity
		CACNA1C cardiomyopathy
		Danon disease
		DES cardiomyopathy
		DSP cardiomyopathy
		Fabry Disease
НСМ		Emery-Dreifuss muscular dystrophy
		FHOD3 cardiomyopathy
		FLNC missense cardiomyopathy
		<i>FLNC</i> tv ACM
LQTS		Histiocytoid cardiomyopath
LVNC		Holt-Oram syndrome
		Tion Oran Synarome
Idiopathic VF		Mitochondrial cardiomyopath PRKAG2 cardiomyopathy
Possible CPVT		
Possible HCM		SCN5A disease
Possible LOTS		Sick sinus syndrome
		TMEM43 cardiomyopathy
Undiagnosed		77Ntv DCM
Clinical Diagnosis	Cardiomyopathies Inherited Arrhythmias	Final Diagnosis



Fig. 5 Diagnostic yield of genetic testing by major ancestry groups. 'Other ancestry' includes Oceanian and People of the Americas. The proportion of suspicious VUS compared between ancestry groups using chi-square. *Abbreviations*: LP/P, likely pathogenic and pathogenic variants; VUS, variant of uncertain significance

clinical geneticist (n = 5, 11%); and referral to a specialised Fabry disease clinic (n = 2, 4%) including one who subsequently commenced enzyme-replacement therapy. One patient found to have Danon disease was expedited for review and subsequent cardiac transplantation [29].

Monogenic disease score

There were 283 (32%) of the total cohort with a positive family history of disease, 274 (31%) were considered to have a severe phenotype, and 244 (27%) diagnosed 30 years of age including 7 (0.8%) diagnosed 10 years of age. These characteristics were used to calculate their likelihood of having a monogenic disease (Additional file 1:

Table S4). Most individuals accumulated few points, with 394 (45%) scoring 0, 162 (18%) scoring 1, 153 (17%) scoring 2, 97 (11%) scoring 3, and 82 (9%) scoring 4. The diagnostic yield increased for those with higher scores, i.e. increasing the likelihood of monogenic disease (Table 2). The yield of LP/P variants ranged from 21% (those with a score of 0) up to 66% (scores 4; Fig. 1A).

Among the 494 individuals with at least one of these characteristics suggestive of monogenic disease, the diagnostic yield of genetic testing was lowest among individuals with only a severe phenotype (LP/P; 22/82, 27%) or only age of disease onset 30 years (LP/P; 24/82, 29%; Fig. 1C), each allocated 1 point under our proposed

	Monogenic disease sco	re			
	0 (likely complex)	1	2	3	\geq 4 (likely monogenic)
Definite clinical diagnosis	362	121	127	85	67
Likely pathogenic/pathogenic	82 (23)	40 (33)	77 (61)	58 (68)	50 (75)
Suspicious VUS	27 (7)	9 (7)	18 (14)	6 (7)	6 (9)
Uncertain clinical diagnosis	32	41	26	12	15
Likely pathogenic/pathogenic	2 (6)	7 (17)	5 (19)	6 (50)	4 (27)
Suspicious VUS	3 (9)	3 (7)	0 (0)	0 (0)	1 (7)
Total cohort	394	162	153	97	82
Likely pathogenic/pathogenic	84 (21)	46 (28)	82 (54)	64 (66)	54 (66)
Suspicious VUS	30 (8)	12 (7)	18 (12)	6 (6)	7 (9)

Table 2 Diagnostic yield of genetic testing

Data are shown as n (%) of each score group

VUS variant of uncertain significance

scoring system. When both of these characteristics were present, the yield increased to 44% (21/47, p < 0.05). The yield of LP/P variants among individuals with only a positive family history (allocated 2 points) was 58% and likewise increased when present in combination with a young age of onset (21/33, 64%), severe phenotype (43/63, 68%), or both (54/82, 66%).

Among those with uncertain clinical diagnoses at baseline evaluation, the overall yield of genetic testing was 24/126 (19%). However, stratifying these cases based on their likelihood of having a monogenic disease was able to improve the genetic yield. For those with a monogenic disease score of 0 (i.e. likely complex disease), there were only 2/32 (6%) individuals with a LP/P variant identified. While those with scores of 3 and 4 had a yield of 6/12 (50%) and 4/15 (27%), respectively (Table 2).

The four highest-scoring families where a genetic diagnosis could be made are shown in Table 3 and include individuals with atypical and severe clinical presentations and causative variants identified via research-based whole exome or genome sequencing in three and comprehensive clinical cardiac panel in one.

Unsolved cases

There were 485 (55%) patients of the total cohort who remained genetically unsolved after both clinical and genetic investigations, of whom 205/485 (42%) were considered unsolved with likely monogenic disease and 280 (58%) unsolved with a likely complex aetiology (Fig. 2).

Individuals without a genetic diagnosis but with clinical features suggestive of monogenic disease (n = 205) scored ≥ 1 and were classified as being likely monogenic. There were 136/205 (66%) with a definite clinical diagnosis at baseline and 69 (34%) with uncertain clinical diagnoses. There were 62 (30%) who had not undergone

expanded genetic testing to date. The highest-scoring unsolved cases are described in Table 3 and include individuals with variable clinical presentations, family history, and research-based whole-exome sequencing so far negative.

Among the 280 considered unsolved with a likely complex disease basis, 27 did not have diagnostic clinical findings at baseline evaluation and were classified as uncertain. No individuals had a severe phenotype and positive family history, and all had age at onset over 30 years. There were 93 individuals who had not undergone expanded genetic testing, and 207 (74%) had a diagnosis of HCM.

Discussion

We report our 18-year experience of genetic testing in a specialised cardiac genetic clinic and demonstrate a role for genetic testing in clarifying the overall diagnosis and patient management. While genetic testing is indicated for those with a definite clinical diagnosis, our dynamic understanding of the phenotypic spectrum of these diseases and the presentation of individuals with undiagnosed or uncertain clinical diagnoses underscores the limitations of this approach. We show that factors indicative of monogenic disease, such as positive family history, severe phenotype, and/or young age at onset, can identify those most likely to achieve a genetic diagnosis. Our findings support genetic testing being offered to all individuals with a confirmed or suspected inherited cardiomyopathy or arrhythmia syndrome and subsequent triaging of cases with characteristics suggestive of a monogenic disease basis for additional, comprehensive research-based sequencing. At present, there are inequities in genetic yield based on ancestry, and this means not all patients will benefit from genetic testing equally.

Table 3 🔾	linical ar	nd genetic features of high scoring solved and unsolve	ed families	
Family code	e Score	Proband	Family history	Genetic testing
Unsolved (c	ongoing	research)		
VE	Ś	Clinically diagnosed with HCM < 10 years with moderate LVH. SCD < 30 years, with HCM identified as the cause of death at post-mortem	Second-degree relative with confirmed ARVC	Negative WES
AMZ	4	SCD < 30 years with moderate hypertrophy and dilata- tion and minor RV fibrosis and fatty infiltration at post- mortem. No clinical diagnosis	Multiple syncopal episodes and RV dilatation in one sibling. Diagnostic type 1 Brugada pattern on ajmaline challenge in multiple relatives.	Negative WGS in the proband and negative WES in the first-, second-, and third-degree relatives
CFR	4	Multiple syncopal episodes, dyspnoea, and 6-s sinus pause on loop recorder onset < 30 years. Mild LV dilata- tion on CMR. No clinical diagnosis	SUD during sleep in parent age < 40 years and one sibling with unclassified sinus node disease and PPM	Negative gene panel and WES
ABM	4	SCA < 30 years, structurally normal heart with atypical ECG changes. ICD implanted for secondary prevention	Children with ECG changes including bradycardia, implanted with PPM	Negative gene panel and WES
Solved on r	eview			
Ξ	4	SCA during pregnancy, apical LVH leading to a clinical diagnosis of HCM (ICD implanted). Later decompensated, progressing to severe heart failure. The final diagnosis of <i>ACTN2</i> cardiomyopathy	Multiple affected relatives with a heterogeneous phenotype ranging from asymptomatic to syncope, heart failure, and SCD	LP variant in ACTN2 identified via research-based linkage analysis
RY	4	Mild apical LVH and atypical ECG changes, clinically diag- nosed with HCM (borderline). The final diagnosis of <i>FHL1</i> cardiomyopathy, with a neurological review identifying elbow contractures and mild muscular dystrophy	SCD in siblings < 20 years	Negative gene panel, LP variant in <i>FHL1</i> identified on WES
D	4	SCD < 30 years, diagnosed with possible HCM at postmortem investigation. The final diagnosis of SCN5A disease	Family members experienced syncope and multiple rela- tives subsequently diagnosed with Brugada syndrome	Negative gene panel, LP translocation in SCN5A on WGS
CPV	m	Elite athlete, presented with SCA < 30 years, but no clini- cal diagnosis despite extensive investigation. The final diagnosis of <i>DSP</i> cardiomyopathy	No family history of the disease	P truncating variant in <i>DSP</i> identified on a comprehensive cardiac panel
SCD sudden o cardiac arrest ventricular hy	cardiac de. <i>, ICD</i> impl: /pertrophy	ath, HCM hypertrophic cardiomyopathy, <i>ARVC</i> arrhythmogenic rig1 antable cardioverter defibrillator, <i>WES</i> whole-exome sequencing, <i>V</i> y	ht ventricular cardiomyopathy, <i>SUD</i> sudden unexpected death w <i>W</i> GS whole genome sequencing, <i>RV</i> right ventricle, <i>CMR</i> cardiac n	ith no cause identified, ECG electrocardiogram, SCA sudden nagnetic resonance, PPM permanent pacemaker, LVH left

Despite decades of research effort, the diagnostic yield of genetic testing for inherited cardiac conditions has barely increased. More stringent variant classification criteria have raised the threshold required to consider a variant; the cause of disease and gene curation efforts attempt to guide consideration of variants only in genes with robust clinical validity for the phenotype in question. It is widely accepted that genetic testing should be offered to patients with inherited cardiac conditions, though there are numerous challenges: access to genetic counselling, which genetic test to order; result interpretation; uncertain findings; secondary findings; and reclassification of variants over time [2, 30]. Despite this, the benefit of a genetic diagnosis for a patient and their family cannot be underestimated. Historically, we have advocated the role of genetic testing in guiding cascade testing of at-risk relatives, allowing us to determine which relatives have the family-specific variant and should continue clinical surveillance. Indeed, the option to release at-risk relatives from ongoing surveillance is the key driver making genetic testing cost-effective compared to clinical screening alone [31-33].

The role of genetic testing in clarifying diagnosis and guiding management is less well described, though studies are now increasingly supporting this approach. For example, genotype-specific evaluation of cases with pathogenic DSP variants allowed recognition of the phenotype as a left-dominant ACM and highlighted that the 2010 Task Force Criteria [34] are inadequate for clinical diagnosis [35, 36]. Limiting access to genetic testing only when clinical diagnostic criteria are met is problematic especially for diseases with poorly defined clinical classifications such as ACM. Rather, an approach that employs key indicators of monogenic disease to better stratify individuals most likely to receive a genetic diagnosis could have greater utility. Positive family history provides high certainty of a shared genetic cause for disease and has been previously associated with a greater diagnostic yield of genetic testing for multiple inherited cardiac conditions [9, 37].

Phenotype extremes and severe outcomes are likewise more often associated with monogenic disease, indeed those with multiple LP/P or VUS in sarcomere genes are known to have more severe diseases [6, 38]. Disease onset at a young age can also indicate a monogenic cause, and in those with severe disease and very young onset, de novo causes are often implicated [39]. Taken together, what we understand about rare genetic variants causing disease could better inform our approach to improving the genetic yield among individuals with inherited cardiac conditions.

For those unlikely to have a strong genetic component to their disease, the contribution of many common variants to disease risk, in combination with environmental factors, is strongly suspected. Polygenic risk scores (PRS), providing a weighting of these common variants, will explain disease for a proportion of patients [4, 5]. In HCM, this is likely to be \sim 40%, including those who have no family history of disease, no LP/P variants, and overall later onset and milder phenotype compared to sarcomere-positive HCM [9]. The overlap between monogenic and polygenic disease has been previously described, including among those with hypercholesterolemia where the highest PRS deciles, inferring the greatest risk, give rise to phenotypes equivalent to those with a monogenic basis [40]. Identifying those with a complex basis for their disease will have marked clinical utility, leading to revised inheritance risks and clinical surveillance advice for relatives. Future research could examine whether risk factors for poor outcomes such as SCD hold true for this group and whether precision therapies currently on the horizon will be equally as effective as those for monogenic disease. Delineating this group can mean we remove them from our 'unsolved' group and better define the diagnostic yield of genetic testing.

Consideration of a complex or monogenic disease basis will allow us to better focus resource-intensive gene discovery efforts. Patients who remain gene elusive following genetic testing and with monogenic disease characteristics are enriched for new insights and discoveries that could have a transformative impact on our understanding of inherited cardiac conditions. A genetic diagnosis can be made in a further 5% of probands through additional research effort. However, the impact of applying more systematic deep phenotyping and cuttingedge genomic analysis tools to this group is currently unknown. The comprehensive, multidisciplinary, and multi-omics approach by undiagnosed disease networks [41] has demonstrated the potential to discover new genes, uncover new disease mechanisms, expand phenotypes of known diseases, and identify new rare diseases. Such discoveries could offer important insights into the underlying biology and potential drug targets, spurring precision management and novel therapies. Solving the cause of disease in these individuals will take bespoke and resource-intensive efforts; therefore, methods to prioritise cases most likely to have an underlying monogenic disease are critical.

Our findings support genetic testing being offered to *all* patients with known or suspected heritable disease, with LP/P variants identified in 21% of those considered to have a likely complex disease basis and suspicious VUS in an additional 8%. We caution that the poor diversity of genetic reference databases critically limits our ability to interpret genetic findings in underrepresented populations. Among our cohort, 15–20% of non-European

background individuals had suspicious VUS identified, compared to only 10% of Europeans. Not only does this translate to fewer patients with a meaningful genetic result, allowing their family members to have cascade genetic testing, but increases the risk of a variant being misclassified with potential for harm [42]. Efforts to increase diversity and be more inclusive in research studies will ensure the discoveries we make apply to the broader community.

Our study has important limitations, including that it was a retrospective audit with a risk of bias in medical record review. The proposed monogenic disease scoring system was developed based on the literature and our clinical experiences, and applying it retrospectively to our cohort supported its clinical value; however, further independent validation is needed. Additionally, the cohort were those referred to a highly specialised adult clinic and may not be representative of the general population; nonetheless, the consecutive patient population makes our data applicable to 'real-world' specialised clinic settings.

Conclusions

We present an 18-year experience of consecutive patients attending a specialised cardiac genetic clinic, highlighting the role of genomics in diagnosis and management. Genetic yield is not equitable across ancestry groups. Factors indicative of monogenic disease can identify those most likely to achieve a genetic diagnosis, even when the clinical phenotype is uncertain. We propose triaging of cases with characteristics suggestive of a monogenic disease basis for additional, comprehensive research-based sequencing.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s13073-022-01149-0.

Additional file 1: Table S1. Tier 1 and Tier 2 gene lists. Table S2. Classified variants. Table S3. Classified suspicious VUS. Table S4. Severe phenotype characteristics used in monogenic disease score. Fig. S1. Reasons for additional genetic diagnoses.

Authors' contributions

FS, NK, ER, and JI designed the study. FS, NK, ER, SH, LY, AB, and JI collected the data. FS, NK, ER, and JI analysed the data. FS, NK, and JI wrote the manuscript. All authors provided critical revision of the manuscript. The authors read and approved the final manuscript.

Authors' information

Twitter handle: @jodieingles27 (Jodie Ingles)

Funding

BG is the recipient of a National Health and Medical Research Council (NHMRC) Early Career Fellowship (#1122330). RS is the recipient of a Vanguard Grant from the National Heart Foundation of Australia (#103082). LY is a recipient of a co-funded National Heart Foundation of Australia/NHMRC PhD scholarship (#102568/ #191351). RB is supported by a New South Wales Health Cardiovascular Disease Senior Scientist Grant. CS is the recipient of a NHMRC Practitioner Fellowship (#1154992) and a New South Wales Health Cardiovascular Disease Clinician Scientist Grant. JI is the recipient of an NHMRC Career Development Fellowship (#1162929). This work was supported in part by a NSW Health Cardiovascular Health Early Mid Career Researcher grant.

Availability of data and materials

Data were collected under a waiver of consent and cannot be deposited. They will be made available upon request and following appropriate ethical approval.

Declarations

Ethics approval and consent to participate

Ethics approval was granted by the Sydney Local Health District Human Research Ethics Committee (X20-0522 and 2020/ETH03142) including a waiver of consent for the collection of de-identified data from all participants attending the specialised clinic.

Consent for publication

Not applicable.

Competing interests

J. Ingles receives research grant support from Bristol Myers Squibb. The remaining authors declare that they have no competing interests.

Author details

¹Cardio Genomics Program at Centenary Institute, The University of Sydney, Sydney, Australia. ²Centre for Population Genomics, Garvan Institute of Medical Research, and UNSW Sydney, 384 Victoria Street, Darlinghurst, NSW 2010, Australia. ³Centre for Population Genomics, Murdoch Children's Research Institute, Melbourne, Australia. ⁴Faculty of Medicine and Health, The University of Sydney, Sydney, Australia. ⁵Agnes Ginges Centre for Molecular Cardiology at Centenary Institute, The University of Sydney, Sydney, Australia. ⁶Department of Cardiology, Royal Prince Alfred Hospital, Sydney, Australia. ⁷Faculty of Medicine, Health and Human Sciences, Macquarie University, Sydney, Australia.

Received: 21 June 2022 Accepted: 12 December 2022 Published online: 28 December 2022

References

- Ackerman MJ, Priori SG, Willems S, et al. HRS/EHRA expert consensus statement on the state of genetic testing for the channelopathies and cardiomyopathies this document was developed as a partnership between the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA). Heart Rhythm. 2011;8:1308–39.
- Hershberger RE, Givertz M, Ho CY, et al. Genetic evaluation of cardiomyopathy - a Heart Failure Society of America practice guideline. J Card Fail. 2018.
- Butters A, Bagnall RD, Ingles J. Revisiting the diagnostic yield of hypertrophic cardiomyopathy genetic testing. Circ Genom Precis Med. 2020;13:e002930.
- Harper AR, Goel A, Grace C, et al. Common genetic variants and modifiable risk factors underpin hypertrophic cardiomyopathy susceptibility and expressivity. Nat Genet. 2021;53:135–42.
- Tadros R, Francis C, Xu X, et al. Shared genetic pathways contribute to risk of hypertrophic and dilated cardiomyopathies with opposite directions of effect. Nat Genet. 2021;53:128–34.
- Ho CY, Day SM, Ashley EA, et al. Genotype and lifetime burden of disease in hypertrophic cardiomyopathy: insights from the Sarcomeric Human Cardiomyopathy Registry (SHaRe). Circulation. 2018;138:1387–98.
- Ingles J, Sarina T, Yeates L, et al. Clinical predictors of genetic testing outcomes in hypertrophic cardiomyopathy. Genet Med. 2013;15:972–7.
- Bos JM, Will ML, Gersh BJ, Kruisselbrink TM, Ommen SR, Ackerman MJ. Characterization of a phenotype-based genetic test prediction score for

unrelated patients with hypertrophic cardiomyopathy. Mayo Clin Proc. 2014;89:727–37.

- Ingles J, Burns C, Bagnall RD, et al. Non-familial hypertrophic cardiomyopathy: prevalence, natural history, and clinical implications. Circ Cardiovasc Genet. 2017:10.
- Barnett IJ, Lee S, Lin X. Detecting rare variant effects using extreme phenotype sampling in sequencing association studies. Genet Epidemiol. 2013;37:142–51.
- Peloso GM, Rader DJ, Gabriel S, Kathiresan S, Daly MJ, Neale BM. Phenotypic extremes in rare variant study designs. Eur J Hum Genet. 2016;24:924–30.
- 12. Maron BJ. Hypertrophic cardiomyopathy centers. Am J Cardiol. 2009;104:1158–9.
- Ommen SR, Mital S, Burke MA, et al. 2020 AHA/ACC guideline for the diagnosis and treatment of patients with hypertrophic cardiomyopathy: a report of the American College of Cardiology/American Heart Association Joint Committee on Clinical Practice Guidelines. J Am Coll Cardiol. 2020;76:e159–240.
- 14. Priori SG, Wilde AA, Horie M, et al. HRS/EHRA/APHRS expert consensus statement on the diagnosis and management of patients with inherited primary arrhythmia syndromes: document endorsed by HRS, EHRA, and APHRS in May 2013 and by ACCF, AHA, PACES, and AEPC in June 2013. Heart Rhythm. 2013;10:1932–63.
- Vohra J, Skinner J, Semsarian C. Cardiac genetic investigation of young sudden unexplained death and resuscitated out of hospital cardiac arrest. Heart Lung Circ. 2011;20:746–50.
- 16. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17:405–24.
- Kelly M, Caleshu C, Morales A, et al. Adaptation and validation of the ACMG/AMP variant classification framework for MYH7-associated inherited cardiomyopathies: recommendations by ClinGen's Inherited Cardiomyopathy Expert Panel. Genetics Med. 2017.
- Whiffin N, Minikel E, Walsh R, et al. Using high-resolution variant frequencies to empower clinical genome interpretation. Genet Med. 2017;19:1151–8.
- Karczewski KJ, Francioli LC, Tiao G, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. Nature. 2020;581:434–43.
- Hosseini SM, Kim R, Udupa S, et al. Reappraisal of reported genes for sudden arrhythmic death. Circulation. 2018;138:1195–205.
- Ingles J, Goldstein J, Thaxton C, et al. Evaluating the clinical validity of hypertrophic cardiomyopathy genes. Circ Genom Precis Med. 2019;12:e002460.
- 22. James CA, Jongbloed JDH, Hershberger RE, et al. International evidence based reappraisal of genes associated with arrhythmogenic right ventricular cardiomyopathy using the Clinical Genome Resource framework. Circ Genom Precis Med. 2021;14:e003273.
- 23. Jordan E, Peterson L, Ai T, et al. Evidence-based assessment of genes in dilated cardiomyopathy. Circulation. 2021;144:7–19.
- Bagnall RD, Ingles J, Dinger ME, et al. Whole genome sequencing improves outcomes of genetic testing in patients with hypertrophic cardiomyopathy. J Am Coll Cardiol. 2018;72:419–29.
- Semsarian C, Ingles J, Bagnall RD. Revisiting genome sequencing data in light of novel disease gene associations. J Am Coll Cardiol. 2019;73:1365–6.
- Chiu C, Bagnall RD, Ingles J, et al. Mutations in alpha-actinin-2 cause hypertrophic cardiomyopathy: a genome-wide analysis. J Am Coll Cardiol. 2010;55:1127–35.
- Yeates L, Ingles J, Gray B, et al. A balanced translocation disrupting SCN5A in a family with Brugada syndrome and sudden cardiac death. Heart Rhythm. 2019;16:231–8.
- Ross SB, Bagnall RD, Yeates L, Sy RW, Semsarian C. Holt-Oram syndrome in two families diagnosed with left ventricular noncompaction and conduction disease. HeartRhythm Case Rep. 2018;4:146–51.
- Martin S, Ingles J, Hunyor I, Bagnall RD, Puranik R, Semsarian C. LAMP2 shines a light on cardiomyopathy in an athlete. HeartRhythm Case Rep. 2017;3:172–6.
- Ingles J, Macciocca I, Morales A, Thomson K. Genetic testing in inherited heart diseases. Heart Lung Circ. 2019.

- Catchpool M, Ramchand J, Martyn M, et al. A cost-effectiveness model of genetic testing and periodical clinical screening for the evaluation of families with dilated cardiomyopathy. Genet Med. 2019.
- Ingles J, McGaughran J, Scuffham PA, Atherton J, Semsarian C. A costeffectiveness model of genetic testing for the evaluation of families with hypertrophic cardiomyopathy. Heart. 2012;98:625–30.
- Wordsworth S, Leal J, Blair E, et al. DNA testing for hypertrophic cardiomyopathy: a cost-effectiveness model. Eur Heart J. 2010;31:926–35.
- Marcus FI, McKenna WJ, Sherrill D, et al. Diagnosis of arrhythmogenic right ventricular cardiomyopathy/dysplasia: proposed modification of the Task Force Criteria. Eur Heart J. 2010;31:806–14.
- 35. Smith ED, Lakdawala NK, Papoutsidakis N, et al. Desmoplakin cardiomyopathy, a fibrotic and inflammatory form of cardiomyopathy distinct from typical dilated or arrhythmogenic right ventricular cardiomyopathy. Circulation. 2020;141:1872–84.
- 36. Hoorntje ET, Burns C, Marsili L et al. Variant location is a novel risk factor for individuals with arrhythmogenic cardiomyopathy due to a desmoplakin (DSP) truncating variant. Circulation: Genomic and Precision Medicine 2022; In press.
- Bos JM, Poley RN, Ny M, et al. Genotype-phenotype relationships involving hypertrophic cardiomyopathy-associated mutations in titin, muscle LIM protein, and telethonin. Mol Genet Metab. 2006;88:78–85.
- Burns C, Bagnall RD, Lam L, Semsarian C, Ingles J. Multiple gene variants in hypertrophic cardiomyopathy in the era of next-generation sequencing. Circ Cardiovasc Genet. 2017:10.
- Acuna-Hidalgo R, Veltman JA, Hoischen A. New insights into the generation and role of de novo mutations in health and disease. Genome Biol. 2016;17:241.
- Natarajan P, Peloso GM, Zekavat SM, et al. Deep-coverage whole genome sequences and blood lipids among 16,324 individuals. Nat Commun. 2018;9:3391.
- Splinter K, Adams DR, Bacino CA, et al. Effect of genetic diagnosis on patients with previously undiagnosed disease. N Engl J Med. 2018;379:2131–9.
- 42. Manrai AK, Funke BH, Rehm HL, et al. Genetic misdiagnoses and the potential for health disparities. N Engl J Med. 2016;375:655–65.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

